

NATURAL DURABILITY OF *Cedrus atlantica* WOOD RELATED TO THE BIOACTIVITY OF ITS ESSENTIAL OIL AGAINST WOOD DECAYING FUNGI

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ABSTRACT

The Atlas cedar, *Cedrus atlantica* is a resinous species of Pinaceae originated from North Africa and well known for its noble timber. This work was conducted to assess the natural durability of its wood, to study the chemical composition of essential oil extracted from its sawdust wood and to test the bioactivity of this essential oil against four wood decaying fungi: *Gloeophyllum trabeum*, *Oligoporus placenta*, *Coniophora puteana* and *Trametes versicolor*. The assessment of natural durability of wood was conducted according to the methods described in the European standards, CEN/TS 15083-1 and NF EN 350-1. Mass losses of wood specimens, after 16 weeks of exposure to fungi attack, in laboratory test, showed that *Cedrus atlantica* wood is very durable to durable against wood decay fungi attack. The extraction of essential oil from sawdust by hydro-distillation yielded about 3,35% and the chemical analysis of this essential oil by GC-MS showed that E- γ -Atlantone (19,73%); E- α -Atlantone (16,86%), 5-Isocedranol (11,68%); 9-iso-Thujopsanone (4,45%); Cedranone (4,13%) and Z α -Atlantone (4,02%) were the main major identified components. The antifungal activity tested by the direct contact technique on agar medium showed a strong inhibition of wood decaying fungi, especially *Gloeophyllum trabeum* inhibited at 1/1000 v/v concentration.

Keywords: Antifungal activity, chemical analysis, essential oils, natural durability, sawdust wood, wood decaying fungi.

INTRODUCTION

The genus *Cedrus*, belonging to the family of Pinaceae, includes four species: *Cedrus atlantica*, *C. libani*, *C. brevifolia* and *C. deodora*. The natural range of *Cedrus atlantica* Manetti is mainly limited to the Algerian and Moroccan mountains (Boudy 1950, Arbez *et al.* 1978). In Morocco, the Atlas cedar covers an area of approximately 132000 hectares located mainly in the Middle Atlas. It annually provides between 80000 and 100000 m³ of wood logs intended for sawing and veneer. This production represents approximately 90% of timber of the total production of the country (HCEFLCD 2013). *C. atlantica* wood is ranked among half-heavy to heavy woods, with basic density, at 12 % moisture, of 530 kg/m³; a total volumetric shrinkage of approximately 11,26%. It has a better dimensional stability and has an easy drying, with axial compression strength of 48,8 MPa, shear strength of 13,6 MPa; a static flexural modulus of 10101 MPa and a rupture under static bending modulus of 94 MPa (El Azzouzi and Keller 1998). Its heartwood is very durable to durable against wood-decay fungi (Brunetti *et al.* 2001).

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Sawing and processing of Cedar wood however generate a lot of waste in form of slabs and sawdust. The last waste form is estimated at about 8% of sawn timber and it represents an important source of essential oils with yields ranging from 2,6 to 5,6% depending on the source and the health of the wood (Aberchane *et al.* 2004, Aberchane *et al.* 2006). Essential oils of cedar, rich in terpenes, were used in perfumery and in the development of many active substances (Adams 1991). Recent studies, on essential oils extracted from cedar wood, highlighted the antimicrobial, antifungal and insecticide effects of these essential oils (Hmamouchi *et al.* 2001, Macchioni *et al.* 2002, Satrani *et al.* 2006, Aberchane 2010, Derwich *et al.* 2010). The use of essential oils extracted from many aromatic and medicinal plants as an agent for wood preservation against wood decay fungi remains little experienced (Haluk and Roussel 2000, El Ajjour *et al.* 2008, Hassane *et al.* 2012).

Relationship between the natural durability of cedar wood and its essential oil bioactivity was not previously related. For this purpose, the present work is to assess the natural durability of *C. atlantica* wood related to the chemical composition and bioactivity of its sawdust wood essential oil against wood decaying fungi with a view to later experiment as a preservative treatment for less durable woods.

MATERIALS AND METHODS

Determination of the natural durability of *Cedrus atlantica* wood

In this study, *C. atlantica* (Manetti ex Endl. Carrière), wood originated from Azrou region (Middle Atlas Mountains, Morocco), was tested and compared to the Scots pine, *Pinus sylvestris* L., sapwood, used as reference wood. Cedar wood specimens of (2,5 cm width, in radial direction \times 1,5 cm thick, in tangential direction \times 5 cm length, in longitudinal direction) were randomly carried from central plates (heartwood) cut in timbers of three trees aged about 90 to 100 years. For biological test, thirty Cedar wood specimens and ten others of Scots pine wood were used for each fungus test. Ten other specimens of these woods were also used for conditioning test. The reference sapwood specimens were carried from untreated commercial Scots pine wood. All specimens were free of cracks, discoloration, biological attack, insect holes and other defects.

Before the test, all specimens were put into a climatic chamber (20 ± 2 °C and $65 \pm 5\%$ relative humidity) in order to reach the wood moisture equilibrium content of 12%.

The three brown rot fungi strains used in this study were *Gloeophyllum trabeum* BAM Ebw.109, *Oligoporus placenta* (ex *Poria placenta*) FPRL. 280, and *Coniophora puteana* BAM Ebw. 15, maintained in the mycological collection of the Laboratory of Botany, Mycology and Environment, Faculty of Sciences, Rabat, Morocco.

The assessment of the natural durability of *C. atlantica* wood is determined according to the European standards: CEN/TS 15083-1 (2005) and NF EN 350-1 (1994). Resistance of woods specimens to fungal decay is based on their mass loss value after fungi exposition in laboratory test compared to those of Scots pine sapwood recognized non-durable against the wood decaying fungi.

The standard CEN/TS 15083-1 previews, before fungi exposition, the calculation of the theoretical oven-dry mass (M_o) of biological test specimens and the K coefficient of moisture correction, obtained from the measurement of the moisture content on another series of ten conditioning specimens of each wood oven dried at 103°C for 24 hours, as indicate:

$$U(\%) = \frac{(M_h - M_o)}{M_o} * 100 \quad (1)$$

$$K = \frac{100}{(100 + U_{\text{moy}}(\%))} \quad (2)$$

where M_h is initial mass at 12%, M_o is anhydrous mass, U (%) is moisture content of each specimen and U_{moy} is mean of moisture content of ten specimens of each type of wood. The theoretical anhydrous mass (M_{to}) of specimens intended for biological tests, is then determined as follows:

$$M_{to} = M_h * K \quad (3)$$

Fungal strains used in this study were grown in Petri dishes on malt-agar medium (4% malt extract and 3% agar in distilled water) and then transferred after 10 to 15 days on the same medium in 500 ml square section bottles. Each bottle, containing 30 ml of the medium, was inoculated with fungi and plugged with cotton. Wood specimens test were at first sterilized by autoclaving at 121 °C for 20 minutes and were exposed to fungal mycelium in bottles after about 20 days of mycelium culture, at the rate of two specimens per bottle. A small round pellets stainless steel of 2 mm thick, used as holders, were placed between the mycelium surface and wood specimens bottom surface. Wood moisture must be above 20 % for suitable wood colonization by the mycelium. Specimens exposed to mycelia were then incubated in a dark climatic chamber (RH= 70 ± 5% and T= 22 ± 2°C) during 16 weeks. At the end of incubation period, the woods specimens were removed from the culture bottles, carefully brushed and immediately weighed to determine their final moisture content before oven dried at 103°C for 24 hours and weighed again to determine their final anhydrous mass (M_f). The mass losses of all inoculated specimens and the average mass loss for each fungus is then determined.

The mass loss, in percentage, of each biological test specimen is calculated as indicate:

$$P = \frac{(M_o - M_f)}{M_o} * 100 \quad (4)$$

M_{to} and M_f are respectively initial theoretical anhydrous mass and final anhydrous mass of wood specimens, and the means of mass loss of the biological test specimens (*C. atlantica* wood, P_e) and the mean of reference wood (Scots pine sapwood, P_r) are calculated as follows:

$$P_{e,r} = \frac{(\sum M_b - \sum M_f)}{\sum M_b} * 100 \quad (5)$$

The durability index “X” of *C. atlantica* wood is therefore:

$$X = \frac{P_e}{P_r} \quad (6)$$

and durability classes of cedar wood are then deducted from the Table 1.

Table 1. Classes of wood durability to brown-rot fungi according to standard NF EN 350-1.

Durability class (DC)	Description	Results expressed in X* value
1	Very durable	$X \leq 0,15$
2	Durable	$0,15 < X \leq 0,30$
3	Moderately durable	$0,30 < X \leq 0,60$
4	Less durable	$0,60 < X \leq 0,90$
5	Non-durable	$X > 0,90$

* X is the durability index expressed as mass loss of the test specimens / mass loss of the reference specimens.

Chemical composition of *Cedrus atlantica* wood essential oil

Three composite samples of sawdust were collected from Cedar wood sawmill of Azrou (Middle Atlas Mountains of Morocco) in heaps of waste on site. The sawdust was then sieved in laboratory to obtain a particle size of 1 mm and prepared for the essential oil extraction.

Three extraction assays of 150 g of sawdust were carried out by hydro-distillation in a Clevenger apparatus for 4 hours. Essential oil obtained was then stored in a small dark glass bottle at 4°C until use. Essential oils yield, expressed in ml/100 g dry matter, was determined related to mean sawdust humidity of three 30 g samples dried at 60°C for 24 hours.

The chemical analysis and components identification were performed by an electronically controlled pressure gas chromatograph (GC) coupled with a mass spectroscope (MS). Gas chromatography analyses were performed with a Hewlett-Packard (HP 6890), equipped with a capillary column HP-5 (30mx0,25mmx0,25µm film thickness) and a detector FID at 250°C. H₂/Air gas mixture was used in split-splitless injector heated at 250°C. The vector gas used was N₂ with 1,5 ml/min. The column temperature was programmed from 50°C to 250°C at 4°C/min. The injected volume of essential oil was 1 µl diluted in n-hexane. A standard solution of n-alkanes (C8-C26) was used to obtain the retention indices. Individual volatile components were identified by comparison of their mass spectra (MS) and retention indices (RI) with those reported in literature and also to the Adams Registry of Mass Spectral Data (Adams 1995).

Bioactivity of *Cedrus atlantica* wood essential oil

The four wood decay basidiomycete fungi used in the biotest were brown wood rots agents (*Gloeophyllum trabeum*, *Oligoporus placenta* and *Coniophora puteana*) and a white rot fungus (*Trametes versicolor*; CTB 863 A strain). They were chosen for the significant damages that they cause to wood and wood-based products.

The antifungal activity of Cedar essential oil was performed by direct contact on agar medium according to the method reported by Remmal *et al.* (1993). In order to give the essential oil a homogeneous distribution in the medium, the oil was first emulsified in a sterile solution of water-agar at 0,2% (SA). To test tubes containing 13,5 ml of malt-agar medium (20 g/l malt extract and 15 g/l agar), sterilized in an autoclave and kept at 45 °C in a water bath, were added aseptically 1,5 ml of different dilutions prepared so as to obtain final dilutions of essential oil in the culture medium of 1/250, 1/400, 1/500, 1/800, 1/1000, 1/1200, 1/2000 and 1/3000. The tubes were shaken vigorously and poured into Petri dishes. Similarly, control plates containing 13,5 ml of culture medium and agar solution at 0,2% (SA) alone were prepared. Petri dishes were inoculated by depositing two square fragments of 0,5 cm², taken from a mycelial mat of a culture of 10 days in malt-agar. Three replicates for each treatment and fungus were prepared and incubated in the dark for 7 days at a temperature of 22 °C. This biotest allows us also to determine the minimum inhibitory concentration (MIC) of essential oil for each fungus tested. The MIC is defined as the lowest concentration for which no growth of the fungus was visually observed (Tantaoui-Elaraki *et al.* 1992).

RESULTS AND DISCUSSION

Natural durability of *Cedrus atlantica* wood

C. atlantica wood specimens showed no apparent decay by the three tested wood decaying fungi and mass losses are below 4,8 % (Table 2). According to our results, this wood is therefore very durable against two fungi, *G. trabeum* and *O. placenta* (durability class, DC1) and durable against *C. puteana* (DC 2). However in view of practical use of the Cedrus wood, further resistance studies should include test fungi with preference for the durable heartwood of pines, like the brown-rot fungi, *Lentinus lepideus*.

The definition of biological risks, according to the standard NF EN 335-2 (2013), is generally taken as a reference in the elaboration of the conditions of end-use of a given wood. A correspondence is then established between natural durability classes and the risk of biological attacks classes (NF EN 460 Standard, 1994). Taking into account this correspondence, natural durability classes (DC 1 and DC 2) of *C. atlantica* wood against wood-destroying fungi, allows this wood to access high-risk classes of biological attacks 4 and 5 for an end-use without preservative treatment, but only regarding decay fungi.

Compared to other coniferous woods, natural durability of *C. atlantica* wood is similar to that of the Atlas Cedar heartwood originated from young Italian plantations, considered as very durable to durable against wood decay fungi (DC 1 and DC 2) (Brunetti *et al.* 2001) and is better than the Aleppo pine timber considered as less durable (DC 4) (Thevenon *et al.* 2012).

Table 2. Mass loss of the test specimens of *C. atlantica* wood and durability classes (DC) according to EN 350-1 in brown-rot test.

	Min %	Max %	Mean %	SD %	X** value	DC***
<i>O. placenta</i>	3,555	6,838	4,745	0,868	0,1437	1
<i>C. puteana</i>	3,230	5,233	4,556	0,468	0,1834	2
<i>G. trabeum</i>	3,163	5,693	4,290	0,622	0,1054	1

(SD standard deviation, n=30, for reference specimens* n=10)

* Means of mass loss of reference specimens (Scots pine sapwood) were respectively 33% for *O. placenta*, 25% for *C. puteana* and 41% for *G. trabeum*,

** Durability index.

*** Durability class.

Chemical composition of *Cedrus atlantica* wood essential oil

The extraction of essential oil by hydro-distillation of Atlas cedar sawdust wood, having a moisture content of about 14,5 %; gave an average yield of essential oil of 3,4%. This is slightly higher than that obtained by the steam stripping distillation (2,4%) reported by Aberchane *et al.* 2001. The yields of Cedar wood essential oil varies greatly depending on the forest source (Aberchane *et al.* 2004), and the part of tree used (Derwich *et al.* 2010, Rhafouri *et al.* 2014). Indeed, distillation needles yielded about 1,8 % while the seeds gave yields between 2,6 and 3,6% (Derwich *et al.* 2010). Low yields (0,05 to 0,49%) were reported by Paoli *et al.* 2011 for essential oils extracted from small branches of *C. atlantica* originated from different localities of Corsica.

Chemical analysis of cedar wood essential oil, by GC-MS permits to identify 41 major components (95,69%) mainly as ketones (52,05%) and alcohols (26,58%) (Table 3). This essential oil is dominated by E- γ -Atlantone (19,73%), E- α - Atlantone (16,86%), 5-Isocedranol (11,68%), 9-iso-Thujopsanone (4,45%), Cedranone (4,13%), Z- α -Atlantone (4,02%), Cedroxyde (2,38%) and 14-Hydroxy- δ -Cadinene (1,94%) (Table 3). This composition is similar to that published by Aberchane *et al.* 2006 using the same extraction method. However, the steam stripping method for the same material revealed that the himachalenes (53%) were major constituents rather than the α -atlantones (14%). This difference in the essential oil composition is due to the extraction method, but also the duration and the distillation temperature can significantly influence the chemical composition of essential oils extracted (Janssen *et al.* 1987, Lachowicz *et al.* 2003).

Furthermore, the essential oil extracted from the needles was dominated by α -pinene (14,85%), γ -himachalene (10,14%) and β -himachalene (9,89%). While the major components of the essential oil of the non-winged seeds were α -pinene (46,16%), manool (25,47%), bornyl acetate (10,18%), β -pinene (5,95%) and α -terpinene (2,71%), those of winged seeds essential oil were manool (49,02%), α -pinene (40,82%), 6-comphenol (2,52%) and β -pinene (2,13%) (Derwich *et al.* 2010, Rhafouri *et al.* 2014). Essential oils extracted from small branches of *C. atlantica* originated from different localities of Corsica were also dominated by α -pinene (up to 79,4%), himachalol (up to 66,2%), β -pinene (up

to 21,49%), β -himachalene (up to 19,3%), γ -himachalene (up to 11,0%), and α -himachalene (up to 10,9%) (Paoli *et al.* 2011).

Table 3. Chemical analysis of *Cedrus atlantica* wood essential oil from Azrou region, the 41 major identified components.

Major Component	RT(mn)	CKI	LKI	PA (%)
Neo-3-Thujanol	15,466	1149,20	1143	1,28
Turmoil	30,267	1579,30	1578	3,44
Carotol	30,715	1593,61	1594	0,21
Cedrol	30,933	1600,61	1596	1,90
Widdrol	31,059	1604,86	1597	0,52
1-Epicubenol	31,291	1612,68	1614	0,40
Himachalol	32,188	1642,93	1647	2,45
α -Cadinol	32,693	1659,97	1653	0,15
5-Isocedranol	32,951	1668,67	1669	11,68
Z-trans Bergamotol	33,523	1687,96	1693	0,45
Kusimol	34,860	1734,56	1736	0,51
β -Santalol	35,081	1742,35	1741	1,98
Z-Epi- β -Santalol	36,859	1805,04	1809	0,49
E-Z-Farnesol	35,306	1750,28	1742	1,12
Total Alcohols				26,58
Hexyl Isobutyrate	15,568	1151,92	1150	1,38
Z-Lingustilide	34,708	1729,20	1730	0,44
Benzyl Benzoate	35,784	1767,14	1762	1,16
E-Ligustilide	36,479	1791,64	1790	0,32
Z- β -Santalol Acetate	37,245	1819,62	1823	1,15
Z-Terpine	37,788	1839,76	1838	1,25
Total Esters				5,69
Camphor	15,466	1149,20	1143	1,28
Cedranone	31,608	1623,37	1620	4,13
9-iso-Thujopsanone	31,867	1632,11	1637	4,45
3-Thujopsanone	32,479	1652,75	1650	0,52
Deodarone	33,235	1678,25	1694	1,07
E γ -Atlantone	34,055	1706,17	1701	19,73
Z α -Atlantone	34,411	1718,72	1713	4,02
E α -Atlantone	36,215	1782,33	1773	16,86
Total Ketones				52,05
Trans Rose Oxyde	14,791	1131,20	1127	0,37
Oxydo himachalene	30,051	1572,40	1574	0,22
β -Himachalene Oxyde	31,166	1608,47	1611	0,41
Cedroxide	34,194	1711,07	1704	2,38
Total oxydes				3,37
Epi-Cedrane	25,956	444,56	1441	0,33
β -Himachalene	27,838	1501,69	1499	0,79
α -Deshydro-ar-Himachlene	28,255	1515,02	1511	1,13
δ -Cadinene	28,545	1524,28	1524	0,31
γ -Deshydro-ar-Himachlene	28,73	1530,19	1529	1,57
α -Calacorene	29,171	1544,28	1542	0,58
β -Calacorene	29,804	1564,50	1563	0,35
14-Hydroxy-Murolene	36,328	1786,32	1775	1,00
14-Hydroxy- δ -Cadinene	36,684	1798,87	1799	1,94
Total Terpenes				7,99
Global				95,69

RT Retention time. LKI Literature Kovàts Index. CKI Calculated Kovàts Index. PA. Peak Area in %.

Bioactivity of *Cedrus atlantica* wood essential oil

The Atlas cedar wood essential oil showed clearly significant antifungal activity against the four wood decaying fungi tested (Table 4). *G. trabeum* fungus was the most sensitive to inhibitory effect of this essential oil since it was inhibited from a concentration of 1/1000 v/v. While a concentration of 1/800 v/v was sufficient to inhibit the growth of *T. versicolor*. However, *C. puteana* and *O. placenta* were the most resistant, their growth inhibition was not noticeable until 1/400 concentrations (Table 4).

Table 4. Antifungal activity of *Cedrus atlantica* sawdust essential oil.

Concentration v/v	1/250	1/400	1/500	1/800	1/1000	1/1200	1/2000	1/3000	C
<i>T. versicolor</i>	-	-	-	MIC	+	+	+	+	+
<i>C. puteana</i>	-	MIC	+	+	+	+	+	+	+
<i>G. trabeum</i>	-	-	-	-	MIC	+	+	+	+
<i>O. placenta</i>	-	MIC	+	+	+	+	+	+	+

C: Control, (-): inhibition, (+): Growth, MIC: Minimal Inhibitory Concentration.

Relationship between natural durability of *Cedrus atlantica* wood and antifungal activity of its essential oil against wood decaying fungi

The antifungal activity of the essential oil of cedar wood can be related to its rich chemical composition essentially atlantones (about 40,61%) which is a sesquiterpen ketones. Studies on the biological activity of some pure compounds of essential oils, such as α -atlantones extracted from *Decalepis hamiltonii*, showed great inhibitory effect especially against pests (George *et al.* 1998) and molds (Thangadurai *et al.* 2002). In our study, alcohols present in significant amount (26,58%), mainly isocedranol, tumerol, himachalol and cedrol (Table 4). May also be involved in the inhibitory effect of this essential oil. The hydrocarbon monoterpenes, such as cadinenes would also have a great antimicrobial property (Keawsa-Ard *et al.* 2012). Furthermore, investigations have already shown that thujaplicines (tropolones) of *Thuja plicata* acted strongly as fungicide substances against Basidiomycetes fungi: *Coniophora puteana*, *Fomes pinicola*, *Lentinus lepideus*, *Serpula lacrymans*, *Polyporus balsameus* and *Poria vaporaria*; and the wood blue stain fungi are also fully inhibited for concentrations in β -thujaplicine of 0,01% (Rennerfelt 1948). The volatile constituents of essential oils such as alcohols create an imbalance in one or the other of the lytic and the synthetic enzymes systems. Growth inhibition therefore follows and it is manifested by stopping natural extension of hyphae (Kahn and Andrawis 1985, Takeuchi and Ichishima 1989).

A synergistic action of two or more components of Cedar wood essential oil can also be involved in the observed bioactivity reported in our study. Furthermore, separate alcoholic and aqueous extracts of *Lipia alba* essential oil, did not have any fungicidal activity at any concentration in culture medium. However *L. alba* essential oil presented fungicide activity against the white-rot fungus, *Pleurotus ostreatus* (Geromini *et al.* 2015).

To identify the active part of Cedar wood essential oil, an antifungal activity of hydrocarbon and oxygenated fractions of this oil is under study.

Bioactivity of essential oil extracted from *C. atlantica* sawdust wood against wood decaying fungi may then explain the level of natural durability of this wood that ranged from very durable (DC1) to durable (DC2). Durability classes were positively correlated with Cedar sawdust essential oil bioactivity against the three wood-destroying fungi specified by the CEN/TS 15083-1 standard (Tables 2 and 4). *G. trabeum* was the most sensitive fungi to this essential oil activity and durability of this wood against the same fungus was the best one.

CONCLUSIONS

The present study, devoted to the assessment of natural durability of *Cedrus atlantica* wood related to the bioactivity of its essential oil, showed that:

The wood of *C. atlantica* is very durable against two fungi. *G. trabeum* and *O. placenta* (durability class. DC1) and durable against *C. puteana* (DC 2).

The essential oil extraction from sawdust yielded about 3,4%; and the composition of this essential oil is dominated by E- γ -Atlantone (19,73%), E- α -Atlantone (16,86%), Isocedranol (11,68%), 9-iso-Thujopsanone (4,45%), Cedranone (4,13%), Z- α -Atlantone (4,02%), Cedroxyde (2,38%) and δ -Cadinene (1,94%).

The essential oil of Atlas cedar sawdust showed clearly significant antifungal activity against the four wood-destroying fungi tested. The high inhibitor oil power might be explained by the high levels of ketones especially γ and α -Atlantones and phenols recognized by their strong fungal inhibitory power.

The durability classes were positively correlated with sawdust essential oil bioactivity against the three wood decaying fungi.

Thus, for best valorization of *C. atlantica* sawdust, its essential oil can be tested in further studies as a wood preservative.

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