

Ions uptake, yield and yield attributes of rapeseed exposed to salinity stress

V. Rameeh^{1*}

¹*Agriculture and Natural Resources Research Center of Mazandran, Sari, Iran. *Corresponding author: vrameeh@yahoo.com*

Abstract

Soil salinity is a serious constrain to crop production in many areas of the world. A pot experiment was carried out with rapeseed genotypes in order to investigate the effects of salinity stress on yield associated traits and shoot ions composition. Eight rapeseed genotypes were evaluated in three salinity levels of irrigation water including 0, 6 and 12 dsm^{-1} . A factorial experiment based on completely randomized design with 4 replications was considered for evaluation of 24 treatments. Significant mean square of the salinity levels, genotypes and salinity \times genotypes interaction effects were exhibited for seeds per pod, pods per plant, seed yield, Ca, K and Na, indicating significant differences of salinity levels and genetic differences for these traits. Due to significant correlation of seed yield with seeds per pod and pods per plant these traits can be considered as indirect selection criteria for seed yield improving. Among shoot ions compositions, K had significant positive correlation with seed yield, therefore this ion can be considered as good indicator for seed yield improving at saline environment. The genotypes including Hyola401, LRT1 and DSM12 had high amounts of seed yield at 12 dsm^{-1} of salinity level and were considered as tolerant genotypes.

Keywords: correlation, experiment, rapeseed, shoot ions, seed yield.

1. Introduction

Salinity stress is a main environmental constraint to crop productivity in the arid and semiarid regions of the world. Soil salinity greatly decreases the productivity of economically important plants including Brassica species. Saline environments influence the plant growth in different methods such as reduction water uptake, an increase of ions to toxic levels, and a reduction of nutrient accessibility. Two clear ways were emphasized for decreasing salinity effects on crop productions: reclamation of salt-affected soils by chemical amendments, and alternatively, the saline soils can be used to grow salt-tolerant plants (Ashraf and McNeilly, 2004). High salt stress also disrupts homeostasis in water potential and ion distribution. This distraction of homeostasis takes place at both the cellular and the whole plant levels (Tunurturk *et al.*, 2011). Tolerance of oilseed brassicas to salt stress is a complex characteristic, which is greatly modified by cultural, climatic and biological factors (Minhas *et al.*, 1990; Kumar, 1995; Mahmoodzadeh, 2008). The amphitetraploids Brassica species including Brassica napus, *B. carinata* and *B. juncea* are more tolerant to salinity and alkalinity than their relevant diploid progenitors such as *B. campestris*, *B. nigra* and *B. oleracea* (Kumar, 1995). Canola (*B. napus* L.) is one of the most important oil seed crops in the world, and even more in Iran, that its production has been notably extended in recent years. A major constraint to seed germination and seedling establishment of canola is soil salinity, which is a common problem in irrigated areas of Iran with low rainfall. This problem adversely affects growth and development of crop, and results into low agricultural production. The most common undesirable effect of salinity on the crop of brassica is the reduction in plant height, size and yield as well as deterioration of the product

quality (Islam, 2001; Zamani *et al.*, 2011). There are differences in sensitivity to salinity among canola cultivars (Puppala *et al.*, 1999; Bybordi, 2010; Tunurturk *et al.*, 2011; Zamani *et al.*, 2011). Calcium (Ca) and potassium (K) ameliorate the adverse effects of salinity on plants (Volkmar, 1998; Munnus, 2002; Amador, 2007). Salinity impairs the uptake of Ca by plants, possibly by displacing it from the cell membrane or in some way affecting membrane function (Lauchli, 1990; Rameeh *et al.*, 2004). Gorham (1993) claimed that all plants discriminate to some extent between sodium (Na) and K. Na can be substituted for K for uptake, and it is believed that similar mechanisms of uptake may operate for both ions (He and Cramer, 1992; Schorder *et al.*, 1994; Porcelli *et al.*, 1995). High levels of K in young expanding tissue is associated with salt tolerance in many plant species (Mer *et al.*, 2000; Ashaf and McNeilly, 2004; Bandeh-Hagh *et al.*, 2008). Closely associated to salt exclusion and its relationship to salt tolerance is the regulation of ion selectivity, in particular the role of Ca/Na and K/Na favoritism in salt tolerance (Sharma and Gill, 1994; Volkmar *et al.*, 1998). Ca could play a regulatory role in the responses of brassica species to saline environments (He and Cramer, 1992). Thakral *et al.* (1998) reported positive non-significant correlation between seed yield and K/Na in stress environment in *B. juncea*. Das *et al.* (1994) claimed that increase in NaCl concentration was associated with increased Na and Cl influx and K efflux in *B. campestris*.

The objective of the present study was to investigate the effect of salinity on yield associated traits and also shoot ions compositions and their relation to seed yield in order to obtain suitable criteria for salinity tolerance in rapeseed genotypes.

2. Materials and methods

In order to study salt stress effect on canola seed yield, yield components and also shoot ions composition, an experiment was conducted in Agriculture and Natural Resources Research Center of Mazandran, Sari, Iran, during 2010-11. The experimental design was a completely randomized design (CRD) arrangement in 3×8 factorial with four replications. Eight diverse rapeseed genotypes (as first factor) including six breeding lines (KRN3, NDK6, KRN1, KRN2, DSM12 and LRT1) and two cultivars (Zarfam and Hyola401) were studied in three salinity levels of irrigation water including 0, 6 and 12 dsm^{-1} as second factor. The salt solution was prepared by taking $\text{NaCl}:\text{CaCl}_2$ in the ratio of 1:1 and the electrical conductivity of different salinity levels was adjusted by a direct reading conductivity meter. Soil analysis results are shown in Table 1. The soil belongs to the non-saline soil with a neutral reaction and the amount of lime which is relatively high. Levels of nutrients, soil organic matter levels in the medium and other nutrients, including potassium, phosphorus, iron, manganese and copper are desirable. In each plot

10 seeds were planted in separate 10-liter pots and five plants were maintained for evaluating. Electrical conductivities of the saline treatments were increased to the desired levels by incremental additions of the salts over 10-day period to avoid osmotic shock to the seedlings. Plants in all pots were irrigated until saturation, with the excess solution allowed to drain into collection pans. All pots were maintained at farm condition and also they were isolated from raining. The studied traits seeds per pod, pods per plant, seed yield, Ca, K and Na. For ions extractions, plant samples were ground by mill and then dried in a furnace at 500°C for 2 hours. After that, plant samples were added 5 mL of 2M HCl for digestion and then they were filtered and diluted by distilled water. The final volume of each sample was 100 mL. Amount of K and Na of each final sample was measured by flame photometer and Ca was measured by atomic absorption (Isaac and Kerber, 1971). Pearson correlation was detected for all the traits. All the traits were analyzed based on factorial experiments based on completely randomized and means comparison were done based on least significant difference test (Gomez and Gomez, 1984).

Table 1. Some of physicochemical properties of soil sample.

Class	(%)			(mg kg ⁻¹)						TNV (%)	OC (%)	pH	Ec (dsm ⁻¹)
	Clay	Silt	Sand	Cu	Zn	Mn	Fe	K	P				
Si-C-L	26	56	18	2.5	0.65	3.2	8	321	7.8	13	1.34	7.2	0.65

3. Results and discussion

3.1. Salinity levels effects on the traits

Significant mean squares of the salinity levels were determined for seeds per pod, pods per plant, seed yield, Ca, K and Na, indicating significant differences of these traits at three salinity levels (Table 2). Seed yield of the genotypes were decreased at high salinity

levels. Pods per plant varied from 22.94 to 13.71 at 0 and 12 dsm^{-1} of salinity levels, respectively (Table 3). The mean of the genotypes was decreased about half its control amount at 12 dsm^{-1} of salinity level. Due to increasing salinity levels, Ca was increased and its amount at 12 dsm^{-1} of salinity level was classified as separate group from the other salinity levels. Since Ca is an integral component of cell wall as well as it is usually bound to the exterior surface of the plasma

membrane, it plays a vital role in maintaining the integrity of both cell wall and membrane. Kopittke *et al.* (2007) was of the view that presence of high quantity of Ca in the cell wall is important so as to provide the plasma membrane with a reasonable amount of Ca to keep its structure intact. Amounts of K were reduced at high salinity levels. The K content in plant tissues represents the main cation in plant cells, and it is a main component of the cell osmotic potential. One of the primary plant responses to salinity is the decrease in K concentration in plant tissues and thus the replacement of K by Na may lead to nutritional imbalances. Both of these ions might compete for entry into

plant root cells. This competition can have significant negative effects on plant growth in saline soils, where concentrations of Na often exceed those of K (Tunurturk *et al.*, 2011). Due to increasing of salinity level, Na was increased and its amount at 12 dsm⁻¹ of salinity level was three times of its control amount. Salinity impairs the uptake up Ca by plants, possibly by displacing it from the cell membrane or in some way affecting membrane function (Lauchli, 1990; Rameeh *et al.*, 2004). Na can be substituted for K for uptake, and it is believed that similar mechanisms of uptake may operate for both ions (He and Cramer, 1992; Schorder *et al.*, 1994; Porcelli *et al.*, 1995).

Table 2. Summary of analysis of variance for yield components, seed yield, Ca, K and Na in rapeseed genotypes.

S.O.V	df	M.S					
		Seeds per pod	Pods per plant	Seed yield (g plot ⁻¹)	Ca (mg g ⁻¹)	K (mg g ⁻¹)	Na (mg g ⁻¹)
Salinity (S)	2	517.52**	709.59**	21.58**	484.00**	1211.33**	1487.06**
Genotypes(G)	7	36.45**	685.72**	7.38**	1322.24**	206.33**	19.30**
S×G	14	12.28**	92.74**	0.43**	278.06**	38.43**	10.17**
Error	72	1.22	6.38	0.11	38.44	10.42	2.22

** : Significant at the 1% levels of probability.

Table 3. Mean comparison of yield components, seed yield, Ca, K and Na for salinity levels based on least significant differences (LSD) test in rapeseed genotypes.

Salinity levels(dsm ⁻¹)	Seeds per pod	Pods per plant	Seed yield (g plot ⁻¹)	Ca (mg g ⁻¹)	K (mg g ⁻¹)	Na (mg g ⁻¹)
S1= 0	13.46a	22.94a	2.84a	28.98b	27.53a	6.327c
S2=6	7.33b	19.94b	1.62b	28.58b	20.86b	13.02b
S3=12	5.89c	13.71c	1.28c	35.51c	15.24c	19.96a

Means, in each column, followed by at least one letter in common are not significantly different at the 1% level of probability.

3.2. Genetic variations of the genotypes

Significant mean square of genotypes at 1% probability level for seeds per pod, pods per plant, seed yield, Ca, K and Na, indicating significant genetic differences of the genotypes for these traits. Seeds per pod ranged from 6.07 to 11.25 related to NDK6 and LRT1, respectively (Table 4). The genotypes including LRNT1, DSM12, Hyola401 and KRN3 with high amounts of seeds per pod were suitable genotypes for improving this trait. The genotypes including Hyola401, LRT and KRN1 had high amounts of pods per plant. Due to significant correlation of seed yield with seeds per pod and pods per plant (0.78** and 0.80**, respectively), these traits can be considered as indirect selection criteria for seed yield improving (Table 6). Seed yield of the genotypes was varied from 0.68 to 3.09 g plot⁻¹ related to Hyola401 and KRN2, respectively. The genotypes including Hyola401, LRT1 and KRN1 with seed yield of 3.09,

2.78 and 2.30 g plot⁻¹, respectively were detected as suitable genotypes for this trait in saline environment. Ca was ranged from 20.19 to 47.77 mg g⁻¹ related to KRN3 and KRN2, respectively. Non significant correlation between Ca and seed yield, indicated that although Ca had ameliorate effects on salinity stress (Volkamar, 1998; Munnus, 2002; Amador, 2007) but in this study it had not important role for seed yield improving of the genotypes. Significant positive correlation was determined between K and seed yield, therefore this ion can be considered as good indicator for seed yield improving at saline environment. Similarly, in earlier studies (Mer *et al.*, 2000; Ashaf and McNeilly, 2004; Bandeh-Hagh *et al.*, 2008) were notified the important effect of K for salinity tolerance. Na was varied from 11.47 to 15.07 mg g⁻¹ related to KRN2 and KRN1, respectively. Due to significant negative correlation between Na and seed yield, the genotypes with low amounts of Na will be preferable for salinity tolerance.

Table 4. Mean comparison of yield components, seed yield, Ca, K and Na for eight rapeseed (*Brassica napus* L.) genotypes based on least significant differences (LSD) test.

Variety	Seeds per pod	Pods per plant	Seed yield (g plot ⁻¹)	Ca (mg g ⁻¹)	K (mg g ⁻¹)	Na (mg g ⁻¹)
1-KRN3	9.63bc	19.00c	1.50d	47.77a	13.07d	13.00bc
2-NDK6	6.87e	12.11d	1.64cd	36.23b	22.11b	11.89c
3-Zarfam	8.33d	13.34d	1.42d	44.59a	22.78b	12.29bc
4-KRN1	8.96cd	23.41b	2.30b	24.4cd	23.06b	15.07a
5-KRN2	6.07e	8.72e	0.68e	20.19d	22.03bc	11.47c
6-DSM12	10.33ab	17.67c	1.89c	23.94cd	27.49a	14.63a
7-LRT1	11.25a	25.22b	2.78a	23.52cd	18.56c	13.59ab
8-Hyola401	9.73bc	31.46a	3.09a	27.54c	20.60bc	12.88bc

Means, in each column, followed by at least one letter in common are not significantly different at the 1% level of probability.

Table 5. Correlation among the studied traits in rapeseed genotypes at different salinity levels.

Traits	Seeds per pod	Pods per plant	Seed yield (g plot ⁻¹)	Ca (mg g ⁻¹)	K (mg g ⁻¹)	Na (mg g ⁻¹)
1- Seeds per pod	1					
2- Pods per plant	0.54**	1				
3- Seed yield	0.78**	0.80**	1			
4- Ca	-0.13	-0.21	-0.24	1		
5-K	0.56**	0.17	0.43*	-0.39	1	
6-Na	-0.73**	-0.27	-0.52**	0.17	-0.69**	1

*, ** Significant at p=5% and 1%, respectively.

3.3. Salinity × genotypes interaction effects on traits

Significant interaction effects of salinity levels × genotypes for all the traits indicated that the trends of variation of these traits among the genotypes were different at the salinity levels. Water availability and nutrient uptake by plant roots is limited because of high osmotic potential and toxicity of Na and Cl ions (Kumar, 1995), therefore due to increasing salinity levels, most of the yield associated traits were decreased. The means of interaction effects of genotypes × salinity levels for the studied traits are presented in Figure 1 to 6 and also in Table 6. Seeds per pod varied from 10.69 to 16 in non saline condition for KRN3 and LRT1, respectively and also it was ranged from 3.23 to 9.45 related to KRN1 and KRN3, respectively at 12 dsm⁻¹ of salinity level (Figure 1). Most of the genotypes with high number of seeds per pod at control condition (0 dsm⁻¹) had high amounts of this trait at the other salinity levels (Table 6). Seeds per pod had less variation for three salinity levels in KRN3. The high amounts of pods per plant were detected for KRN1, LRT1 and Hyola401 at 12 dsm⁻¹ of salinity level (Figure 2), therefore these genotypes can be considered for improving this trait at saline condition. The highest seed yield at control and

12 dsm⁻¹ of salinity level was belonged to Hyola401 and followed by LRT1 and DSM12 had high amounts of this trait at 12 dsm⁻¹ of salinity level (Figure 3). Due to salinity increasing, Ca was increased in most of the genotypes and KRN3, Hyola401 and NDK6 had high amount of Ca at high salinity levels (Figure 4). The concentration of K reduced in the most of the genotypes at high salinity levels and the genotypes including DSM12, NDK6 and Zarfam had high means value of K at highest salinity level (Figure 5). Na was increased in all of the genotypes at high salinity levels and the genotypes including NDK6, Zarfam and KRN2 with low amounts of Na at 12 dsm⁻¹ of salinity level were detected as suitable genotypes. Agricultural productivity is severely affected by soil salinity, and the damaging effects of salt accumulation in agricultural soils have influenced ancient and modern civilizations. The detrimental effects of salt on plants are a consequence of both a water deficit that results from the relatively high solute concentrations in the soil and a Na specific stress resulting from altered K/Na ratios and Na ion concentrations that are inimical to plants. The alteration of ion ratios in the plant is caused by the influx of Na through pathways that function in the acquisition of K (Rameeh et al., 2004; Tunuturk et al., 2011).

Table 6. Mean comparison of yield components, seed yield, Ca, K and Na for eight rapeseed (*Brassica napus* L.) genotypes based on least significant differences (LSD) test.

Variety	Seeds per pod	Pods per plant	Seed yield (g plot ⁻¹)	Ca (mg g ⁻¹)	K (mg g ⁻¹)	Na (mg g ⁻¹)
1-S0 × KRN3	10.69de	26.92bc	2.45cd	45.50abc	15.88h-k	8.63hi
2- S0 × NDK6	12.12cd	21.44e	2.57cd	42.36bc	27.89bc	6.04ij
3- S0 × Zarfam	13.82bc	22.85cde	2.43cd	52.63ab	28.08bc	5.99ij
4- S0 × KRN1	13.06c	21.70de	3.25	16.92fg	34.29a	6.14ij
5- S0 × KRN2	10.66def	11.40f	1.33b	17.88efg	32.78ab	6.11ij
6- S0 × DSM12	15.45ab	13.59f	2.58cd	19.36efg	32.85ab	6.66ij
7- S0 × LRT1	16.00a	28.37b	4.01a	23.01efg	23.09c-g	5.80j
8- S0 × Hyola401	15.91a	37.28a	4.14a	14.15g	25.40cde	5.26j
9-S1 × KRN3	8.75efg	18.96e	1.08ghi	42.45bc	16.20h-k	11.72fg
10- S1 × NDK6	4.86hij	9.85fg	1.40fg	22.27efg	19.83e-j	12.17efg
11- S1 × Zarfam	6.24hi	10.90fg	1.06ghi	38.79cd	22.73c-g	13.36ef
12- S1 × KRN1	10.18def	26.40bcd	2.24de	27.81def	20.37d-i	14.58e
13- S1 × KRN2	4.32ij	10.12fg	0.47ij	24.16efg	22.35c-g	10.48gh
14- S1 × DSM12	8.78efg	20.74e	1.33fgh	27.98def	25.91cd	14.56e
15- S1 × LRT1	8.62fg	26.28bcd	2.87bc	23.48efg	18.51-j	13.79ef
16- S1 × Hyola401	6.91gh	36.31a	2.49cd	21.73efg	21.00d-h	13.50ef
17-S2 × KRN3	9.45ef	11.12f	0.99ghi	55.36a	7.15l	18.65cd
18- S2 × NDK6	3.62j	5.04h	0.96ghi	44.04abc	18.59f-j	17.48d
19- S2 × Zarfam	4.93hij	6.29gh	0.76hij	42.34bc	17.53g-j	17.52d
20- S2 × KRN1	3.65j	22.12de	1.42fg	28.47de	14.53ijk	24.48a
21- S2 × KRN2	3.23j	4.65h	0.26j	18.53efg	10.96kl	17.83d
22- S2 × DSM12	6.75gh	18.69e	1.76ef	24.50efg	23.70c-f	22.67ab
23- S2 × LRT1	9.13ef	21.00e	1.48fg	24.09efg	14.06jk	21.17bc
24- S2 × Hyola401	6.37hi	20.78e	2.66bcd	46.74abc	15.41h-k	19.88cd

Means, in each column, followed by at least one letter in common are not significantly different at the 1% level of probability.

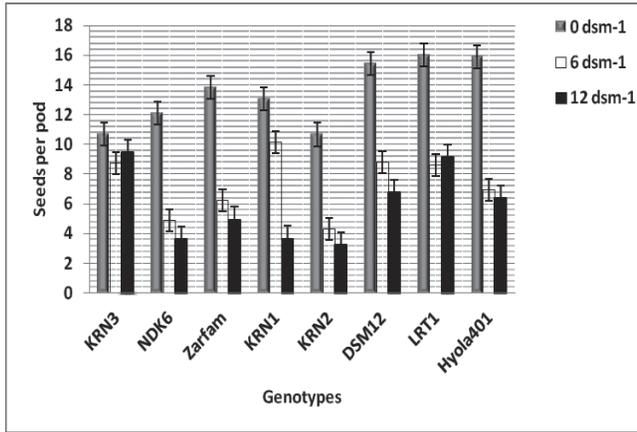


Figure 1. Seed per pod in eight rapeseed genotypes at the different salinity levels.

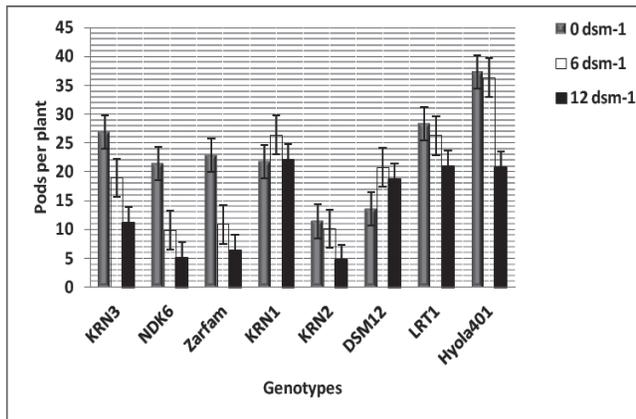


Figure 2. Pods per plant in the eight rapeseed genotypes at the different salinity levels.

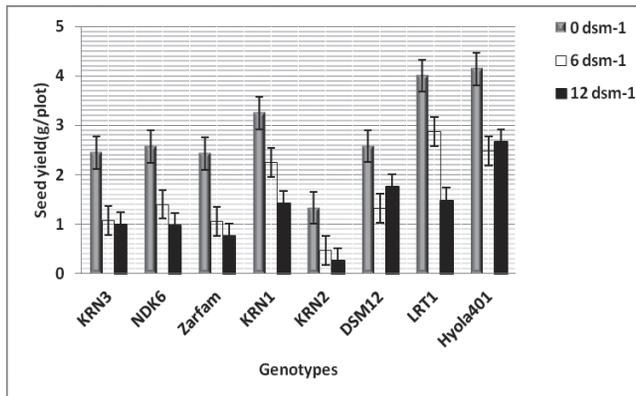


Figure 3. Seed yield of the eight rapeseed genotypes at the different salinity levels.

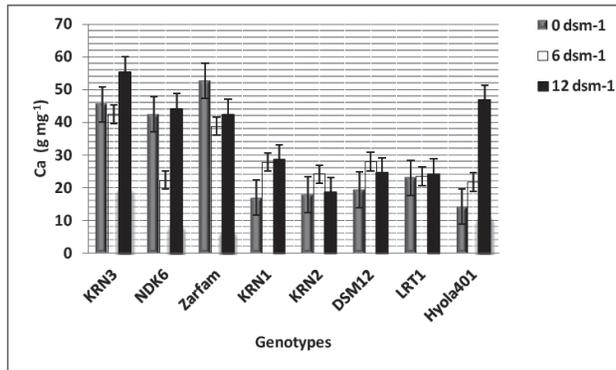


Figure 4. Ca in eight rapeseed genotypes at the different salinity levels.

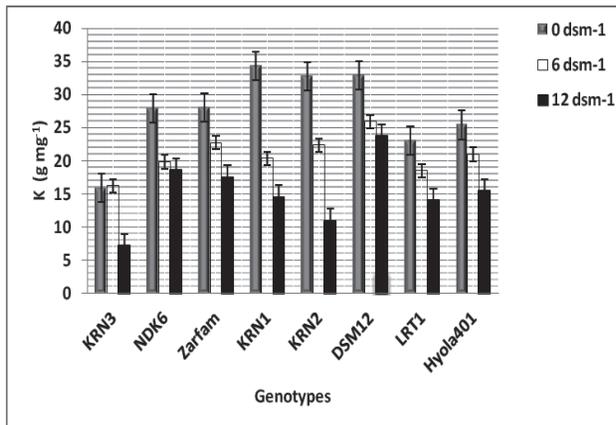


Figure 5. K in eight rapeseed genotypes at the different salinity levels.

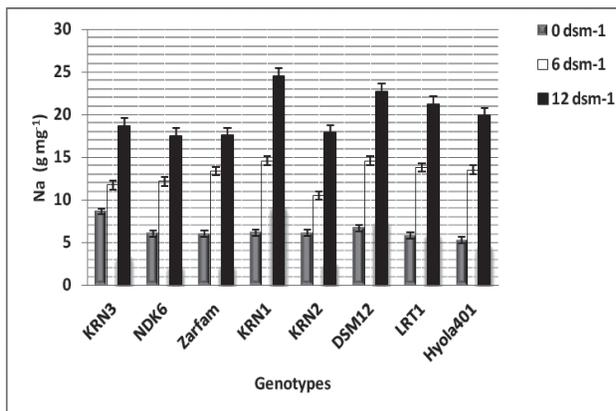


Figure 6. Na in eight rapeseed genotypes at the different salinity levels.

In general salinity levels had significant effects on the traits and due to increasing of salinity levels, yield and yield components and also K were decreased. With increasing salinity levels, Ca also had lowest variations among the genotypes and Na was increased in all of the genotypes. Significant correlation of seed yield with seeds per pod and pods per plant indicating these traits can be considered as indirect selection criteria for seed yield improving. Significant positive correlation was also exhibited between K and seed yield, therefore this shoot ion can be considered as good indicator for seed yield improving at saline environment. Due to existing significant negative correlation between Na and seed yield, the genotypes with low amounts of Na at saline condition will be preferable.

References

- Amador, B.M., Yamada, S., Yamaguchi, T., Puente, E.R., Serrano, N.A.V., Hernandez, J.L.G., Aguilar, R.L., Dieguez, E.T., Garibay, A.N. 2007. Influence of calcium silicate on growth, physiological parameters and mineral nutrition in two legume species under salt stress. *J. of Agron. and Crop Sci.* 193, 413-421.
- Ashraf, M., McNeilly, T. 2004. Salinity tolerance in *brassica* oilseeds. *Critical Rev. of Plant Sci.* 23, 2, 157-174.
- Bandeh-Hagh, A., Toorchi, M., Mohammadi, A., Chaparzadeh, N., Salekdeh, G.H., Kazemnia, H. 2008. Growth and osmotic adjustment of canola genotypes in response to salinity. *J. of Food, Agric. and Environ.* 6,2, 201-208.
- Bybord, A. 2010. Effects of salinity on yield and component characters in canola (*Brassica napus* L.) cultivars. *Not. Sci. Biol.* 2, 1, 81-83.
- Gorham, J. 1993. Genetics and physiology of enhanced K/Na discrimination. In: Genetic aspects of plant mineral nutrition. Randall, P., Ed.; Kluwer Academic Publishers, Dordrecht, The Netherlands. pp:151-159.
- Gomez, K.A., Gomez, A. A. 1984. Statistical procedures for agricultural research; John Wiley and Sons, New York.
- He, T., Cramer, G.R. 1992. Growth and mineral nutrition of six rapid-cycling *Brassica* species in response to seawater salinity. *Plant Soil.* 139,2, 285-294.
- Isaac, R.A., Kerber, J.D. 1971. Atomic absorption and flame photometry: Techniques and uses in soil, plant and water analysis. In: Instrumental Methods for Analysis of Soil and Plant Tissue. Walsh, L. M., Ed.; Soil Sci. Of Am. Madison, Wis. Pp: 17-37.
- Islam, M.R., Bhuiyan, M.A.R., Prasad, B., Quddus, M.A. 2001. Salinity effect on yield and component characters in rapeseed and mustard varieties. *J. of Biol. Sci.* 1,9,840-842.
- Kumar, D. 1995. Salt tolerance in oilseed brassicas-present status and future prospects. *Plant Breed. Abst.* 65, 10,1439-1477.
- Lauchli, A. 1990. Calcium, salinity and the plasma membrane. In: R.T. Leonard and P.K. Hepler (ed.), Calcium in Plant Growth and Development. American Society of Plant Physiologists, Rockville, Maryland, pp. 26-35.
- Mandhan, S., Madan, S., Sheokand, S. 2010. Differential response in salt tolerant and sensitive genotypes of wheat in terms of ascorbate, carotenoids proline and plant water relations. *Asian J. Exp. Sci.* 1,4,792-797.
- Mer, R.K., Prajith, P.K., Pandya, D.H., Pandey, A.N. 2000. Effect of salts on germination of seeds and growth of young plants of *Hordeum vulgare*, *Triticum aestivum*, *Cicer arietinum* and *Brassica juncea*. *J. of Agron. and Crop Sci.* 185, 209-217.

- Minhas, P.S., Sharam, D.R., Khosla, B.K. 1990. Effect of alleviation of salinity stress at different growth stages of Indian mustard (*Brassica juncea*). Indian J. of Agric. Sci. 60, 5, 343-346.
- Munns, R. 2002. Comparative physiology of salt and water stress. Plant Cell Environ. 25, 239-250.
- Naseri, R., Mirzaei, A., Tahereh Emami, T., Vafa, P. 2012. Effect of salinity on germination stage of rapeseed cultivars (*Brassica napus* L.). Intl. J. Agr. Crop Sci. 4,13, 918-922.
- Porcelli, C.A., Gutierrez-Boem, F.H., Lavado, R.S. 1995. The K/Na and Ca/Na ratios and rapeseed yield under soil salinity or sodicity. Plant Soil. 175, 2, 251-255.
- Puppala, N.J., Fowler, L., Poindexter, L., Bhadwaj, H.L. 1999. Evaluation of salinity tolerance of canola germination, In: Perspectives on new crops and new uses. Janick J., Ed.; ASHS press, Alexandria, VA. 251-253.
- Rameeh, V., Rezai, A., Saeidi, G. 2004. Study of salinity tolerance in rapeseed. Commun. Soil Sci. Plant Analysis. 35, 2849-2866.
- Rameeh, V., Cherati, A., Abbaszadeh, A. 2012. Relationship Between Seed Yield and Shoot Ions at Vegetative and Reproductive Stages of Rapeseed Genotypes under Saline. International J. of Plant Res. 2,3, 61-64.
- Schorder, J.I., Ward, J.M., Gassmann, W. 1994. Perspectives on the physiology and structure of inward –rectifying K channels in higher plants: biophysical implications for K uptake. *Annual Review of Biophysics and Biomolecular Structure*. 23, 441-471.
- Sharma, P.C., Gill, S.K. 1994. Salinity-induced effect on biomass, yield, yield-attributing characters and ionic contents in genotypes of Indian mustard (*Brassica juncea*). Indian J. of Agric. Sci. 64,11,785-788.
- Thakral, N., Singh, H., Kumar, P., Yavada, T.P., Mehta, S.L. 1998. Association analysis between physio-chemical parameters with seed yield in Indian mustard under normal and saline environments. Crucifereae Newsletter. 20, 59-60.
- Tunuturk, M., Tuncturk, R., Yildirim, B., Ciftci, V. 2011. Effect of salinity stress on plant fresh weight and nutrient composition of some Canola (*Brassica napus* L.) cultivars. African Journal of Biotechnology. 10, 10,1827-1832.
- Volkamar, K.M., Hu, Y., Steppuhn, H. 1998. Physiological responses of plants to salinity: A review. *Can. J. of Plant Sci.* 78, 19-27.
- Zamani, Z., Nezami, M.T., Habibi, D., Khorshidi, M.B. 2010. Effect of quantitative and qualitative performance of four canola cultivars (*Brassica napus* L.) to salinity conditions. Adv. in Environ. Biol., 4, 3, 422-427.

