

Histomorphometric Evaluation of the Effects of Mandibular Advancement Appliance and Low Level Laser Therapy (LLLT) with Different Doses on Condylar Cartilage and Subchondral Bone in Rats

Evaluación Histomorfométrica de los Efectos del Aparato de Avance Mandibular y la Terapia con Láser de Bajo Nivel (TLBN) con Diferentes Dosis en Cartílago Condilar y Hueso Subcondral en Ratas

Rıdvan Oksayan¹; Oral Sökücü² & Neslihan Üçüncü³

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SUMMARY: The aim of this study was to evaluate the effects of mandibular advancement appliance and low level laser therapy (LLLT) with different doses on cellular hypertrophic changes in the mandibular condyle of rats. Forty-eight 8-week-old male Wistar albino rats weighing between 260 and 280 g were randomly divided into four experimental and control groups. Group I was the control group; group II was the mandibular advancement appliance group; group III was the 8 J/cm² (0.25 W, 20 s) laser irradiation with mandibular advancement appliance group; and group IV was the 10 J/cm² (0.25 W, 25 s) laser irradiation with mandibular advancement appliance group. Mandibular condyle cartilage and subchondral bone changes with different LLLT dose and mandibular advancement appliance were evaluated by histomorphometrical analysis. Subchondral bone fraction results showed that there were no significant differences between groups ($p < 0.05$). The statistically significant differences found between control group and experimental groups in anterior and posterior cartilage layers thickness ($p < 0.05$) and ($p < 0.01$). Posterior and anterior condylar cartilage layers of rats react differentially to LLLT and mandibular advancement application. Maximum changes in condylar cartilage layers were found in 8 J/cm² laser irradiation with mandibular appliance group.

KEY WORDS: Low level laser therapy; Mandibular advancement; Rat; Histomorphometric.

INTRODUCTION

Mandibular deficiency-induced skeletal class II malocclusion has been a common problem in orthodontic practice. Patients with skeletal class II malocclusion and mandibular retrognathia have severe esthetic, psychological, airway and occlusal problems (El-Bialy *et al.* 2003, 2015). Orthopedic therapy in growing patients has the power to treat underdevelop mandibula by enhancing condylar growth. The functional orthopedic treatment of skeletal Class II malocclusion stimulates the upward and backward growth of mandibular condyle, and this process plays an important role in the forward movement of the mandible (Oksayan *et al.*, 2015). As a result, adaptive mandibular condylar cartilage remodelling occurs with the forward movement of mandible by functional appliances (Owtad *et al.*, 2011). According to patients' age, sex, severity of skeletal problem and

compliance, functional appliance therapy is to achieve the condylar cellular activity and bone remodelling in a shorter treatment time.

Many studies have shown that biomechanical stimulus are important for development and growth process of secondary cartilages like mandibular condyle cartilage (Tingey & Shapiro, 1982; Peltomäki *et al.*, 1997). Through the development of new methods and technologies some researchers studied to increase the chondroblastic and osteoblastic activity in mandibular condylar cartilage with laser and ultrasound for mandibular condyle growth stimulation and acceleration with, or without functional appliances in experimental animals (Oyonarte *et al.*, 2009; Seifi *et al.*, 2010; El-Bialy *et al.*, 2015). Low level laser

¹ Department of Orthodontics, Faculty of Dentistry, Eskisehir Osmangazi University, Eskisehir, Turkey.

² Private Practice, Gaziantep, Turkey.

³ Department of Orthodontics, Faculty of Dentistry, Gazi University, Ankara, Turkey.

therapy (LLLT) is gaining popularity in medical and dental fields in last years. In recent experimental animal studies, LLLT has become a foremost way to accelerate and stimulate the condylar cartilage growth and mandibular advancement with or without functional appliance (Seifi *et al.*; Abtahi *et al.*, 2012; El-Bialy *et al.*, 2015).

In this study we aimed to evaluate the effects of mandibular advancement appliance and LLLT with different doses on condylar cartilage layer thickness, total cartilage thickness and subchondral bone changes in anterior and posterior region of mandibular condyle of rats.

MATERIAL AND METHOD

Forty-eight male 8-week-old Wistar Albino rats weighing between 260 and 280 g, divided randomly into four groups. Group I (n=12) was the control group; group II (n=12) was the mandibular advancement appliance group; group III (n=12) was the 8 J laser irradiation with mandibular advancement appliance group; and group IV (n=12) was the 10 J laser irradiation with mandibular advancement appliance group (Table I). The study was approved by İnönü University Experimental Animal Ethical Committee with 2013/A-38 code. The rats were caged in a 23 °C temperature, 12 h night / 12 h day environment and regulated humidity conditions. Animals were fed ad libitum with a standard soft diet to prevent any damage to the mandibular advancement appliances. Experimental rats were stimulated with a low-level laser in the temporomandibular joint region bilaterally 15 times over 30 days for symmetrical growth of mandibular condyle. Histomorphometrical changes in the mandibular condyle cartilage and subchondral bone were evaluated on day 30. The animals were anesthetized with an intramuscular injection of 20 % Ketamine hydrochlorur (Ketalar-Eczacıbaşı/Türkiye) and 80 % Xylazine (Rompun-Bayer/Germany) before the mandibular advancement appliance applications.

Functional mandibular advancement appliance application. The functional mandibular advancement appliance applied in this research was similar to that developed for growing rats in previous studies (Xiong *et al.*, 2004; Owtad *et al.*). Silicon impressions were obtained from mandibular

incisors and plaster models were made. Mandibular advancement appliances were constructed from hard Essix material for the mandibular incisors and identical inclination planes were fitted to the lower incisors of rats in the experimental groups. And mandibular advancement appliances repositioned the rats mouths vertically open and cross bite position. These appliances bonded to lower incisors with the self etch bonding system (Transbond Plus, 3M, Monrovia, USA) and advancement appliances provided 2.5–3 mm continuous forward mandibular movement in experimental groups (Fig. 1). The functional mandibular advancement appliance was not applied to rats in the control group.

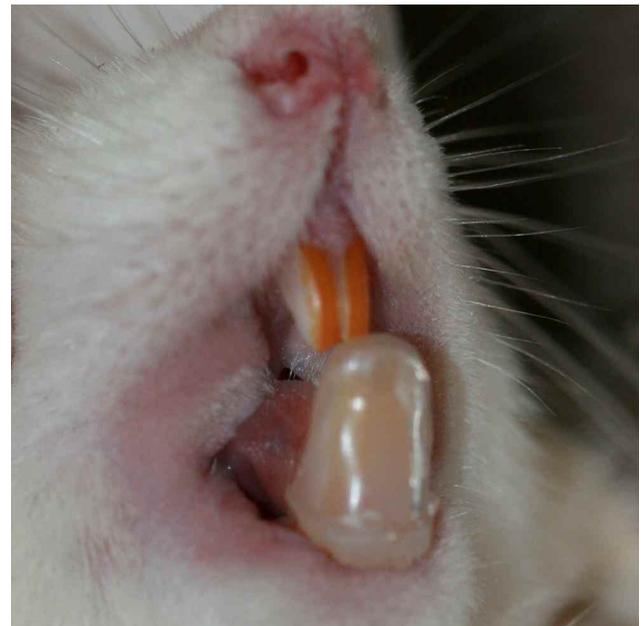


Fig. 1. Experimental mandibular advancement appliance for rat mandibular incisors.

Laser application. Laser applications were performed in temporomandibular joint region of rats in group III and IV with a 810 nm GaAs diode laser (Cheese, Wuhan, China) with a probe diameter of 0.625 cm². Application was applied bilaterally and in direct contact to the skin 15 times in 30 days. In group III received 8 J/cm² (0.25 W, 20 sec) energy density on each side per session. And group IV received 10 J/cm² (0.25 W, 25 sec) energy density on each side per session.

Histomorphometrical preparations and analysis. All experimental and control group rats were sacrificed with injection of high amount of anesthesia. Left condyles of rats were dissected and suspended in 10 % formalin solution (Merck, Darmstadt, Germany) in 48 hours. After this protocol, condyle specimens were decalcified in 5 % formic acid solution (Tekkim, Bursa, Türkiye) for 2 weeks at 4 °C.

Table I. Design of experimental study protocol.

Groups	N	Appliance	LLLT energy density	Experimental period time
Grup I	12	-	-	30 days
Grup II	12	+	-	30 days
Grup III	12	+	8 J/cm ²	30 days
Grup IV	12	+	10 J/cm ²	30 days

Samples washed with distilled water at the end of the 2 week and tissue was placed in the trays. Rapid biopsy program was started with the help of automatic tissue processing machine (Thermoscientific, ExcelsiorES, USA) to follow tissue. Then the samples were embedded in paraffin blocks (Gurr, Leuven, Belgium). For examination of cartilage and subchondral structures clearly, outer surface of the condyles were trimmed 200-250 μm . Parasagittal serial sections (5 μm) were cut and stored at 37 °C in the oven for approximately 24 h. Hematoxylin eosin (Surgipath, Peterborough/USA) staining procedure were applied to each sample for histomorphometrical evaluations. The samples obtained from subjects were examined under light microscope (Leica DM5000 B, Wetzlar/Germany) and Image

acquisition was performed Leica Qwin.plus image analysis system (Leica Qwin.Plus, Cambridge, United Kingdom).

The condylar cartilage was equally divided into six regions (three of them anteriorly and three of them posteriorly) and the condylar cartilage was typically organized into the fibrous, proliferative, mature, and hypertrophic layers (Jiao *et al.*, 2010; Li *et al.*, 2013) (Fig. 2). Thickness of condylar cartilage layers and total layers were measured at the six regions (three of them anteriorly and three of them posteriorly) of the condylar cartilage. The three measurements in anterior and posterior parts were averaged. In the middle of the anterior and posterior parts of the condyle two square (0.5 mm \times 0.5 mm) bone area fraction (bony area / total area) measurement was performed under the interface of hypertrophic cartilage layer (Jiao *et al.*) (Fig. 3).

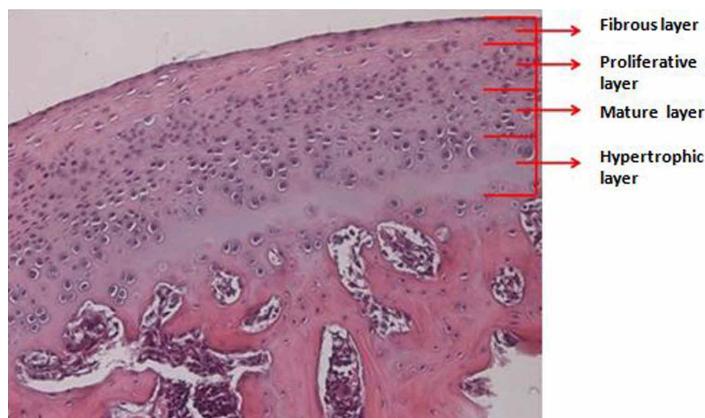


Fig. 2. Hematoxylin eosin staining of rat condylar cartilage layers (fibrous, proliferative, mature, and hypertrophic) (200x).

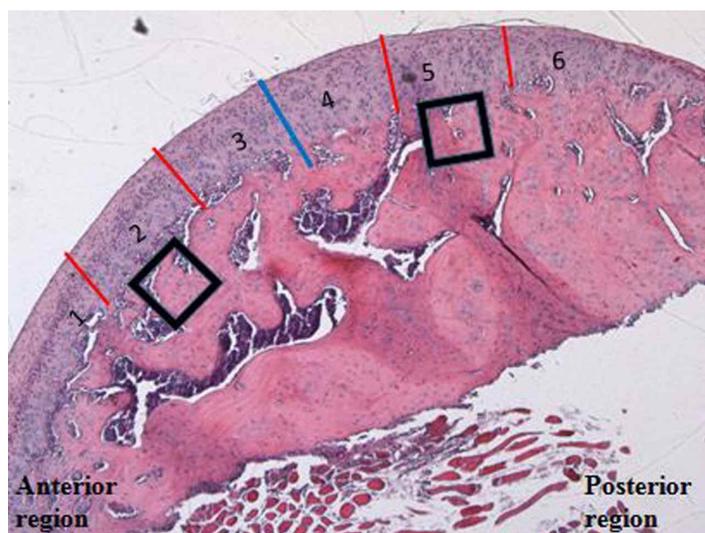


Fig. 3. Figure of condylar cartilage layers and the subchondral bone area measurements. Thickness of condylar cartilage and total layers were examined at the six regions (three of them anteriorly and three of them posteriorly). In middle of the anterior and posterior parts of the condyle two square (0.5 mm \times 0.5 mm) bone area fraction (bony area / total area) measurement was performed under the interface of hypertrophic cartilage layer (50x).

Statistical analysis. Statistical analysis was performed using SPSS software (Statistical Package for Social Science, SPSS Inc. Chicago, Illinois, America, Windows version 20.0). To evaluate the normality Kolmogorov-Smirnov test was used. The nonparametrical statistical Kruskal-Wallis and Mann-Whitney U tests were applied to analyze the intragroup and intergroup significant differences on histomorphometric datas.

RESULTS

Body weight. Body weight of the animals did not change in 30-day experimental process in any group. The increase of the weight in the group I (control) was more than that of other experimental groups (Table II).

Subchondral bone. According to subchondral bone area fraction (bony area / total area) results, anterior and posterior subchondral bone measurements demonstrated no significant differences between the groups ($p < 0.05$) (Table III).

Anterior condylar cartilage layers. The results of this research showed that anterior hypertrophic layer thickness in condylar cartilage was not significantly changed, with LLLT and a mandibular advancement device during the 30 day experimental period ($p < 0.05$).

For anterior mature layer thickness parameter, a statistically significant difference was

detected between the groups ($p < 0.01$). Group III ($p < 0.01$) and IV ($p < 0.05$) showed statistically significant low thickness values than group I (control).

A statistically significant difference was found between the groups for anterior proliferative layer thickness in condylar cartilage ($p < 0.01$). Experimental groups III ($p < 0.01$) and IV ($p < 0.05$) showed statistically significant low thickness values according to group I (control).

According results of this experimental study anterior fibrous layer thickness in condylar cartilage was not significantly changed with LLLT and mandibular advancement during the experimental period ($p < 0.05$) (Table IV).

Posterior condylar cartilage layers. Histomorphometrical results for posterior hypertrophic layer thickness in condylar cartilage showed statistically significant differences between the groups ($p < 0.05$). Group II ($p < 0.01$), group III ($p < 0.05$) and IV ($p < 0.01$) showed statistically significant low thickness values than group I (control).

Statistically significant differences were found between groups in posterior mature layer thickness of condylar cartilage variable ($p < 0.05$). Group II ($p < 0.05$), group III ($p < 0.01$) and IV ($p < 0.01$) showed statistically significant low thickness values than group I (control).

For posterior proliferative layer thickness parameter, a statistically significant difference was detected between the groups ($p < 0.05$). Group II ($p < 0.05$), group III ($p < 0.05$) and IV ($p < 0.01$) showed statistically significant low thickness values than group I (control).

According results of this experimental study posterior fibrous layer thickness in condylar cartilage did not showed statistically significant differences between groups with LLLT and mandibular advancement during the experimental period ($p < 0.05$) (Table V).

Table II. Body weight of rats before and after the experimental study period.

Variables	N	Groups	Mean	Standart	P
Anterior subchondral bone area fraction (bony area / total area) (%)	1	I	0.55	0.13	0.115
	1	II	0.44	0.13	
	1	III	0.45	0.12	
	2	IV	0.41	0.11	
Posterior subchondral bone area fraction (bony area / total area) (%)	1	I	0.36	0.16	0.369
	1	II	0.42	0.17	
	1	III	0.43	0.15	
	2	IV	0.34	0.16	

Table III. Comparisons of subchondral bone area fraction (bony area / total area)(%) measurements between groups. ($p < 0.05$)

Groups	T ₀ (g)	T ₁ (g)
Group I	262.75 ± 48.69	281,5 ± 50.81
Group II	268.7 ± 22.54	272.1 ± 22.27
Group III	273.81 ± 25.51	275.27 ± 21.77
Group IV	270.75 ± 30.98	272 ± 33.23

Table IV. Statistical results of anterior layer thickness in condylar cartilage according to Kruskal-Wallis and Mann Whitney U tests.

Variables	Group I Mean ±S.D.	Group II Mean ± S.D.	Group III Mean ± S.D.	Group IV Mean ± S.D.	P	Multiple comparisons between groups				
						I-II	I-III	I-IV	II-III	
Anterior hypertrophic layer (µm)	162.08 ± 148.52	73.56 ± 46.84	75.57 ± 49.52	112.11 ± 105.51	0.059	NS	NS	NS	NS	NS
Anterior mature layer (µm)	61.48 ± 25.85	40.96 ± 34.10	22.81 ± 8.66	40.51 ± 26.17	0.003**	NS	0.000**	0.015*	NS	NS
Anterior proliferative layer (µm)	47.33 ± 19.58	33.17 ± 27.87	19.79 ± 7.67	33.92 ± 23.60	0.008**	NS	0.000**	0.035*	NS	NS
Anterior fibrous layer (µm)	71.28 ± 60.32	64.42 ± 88.37	28.22 ± 23.42	56.83 ± 42.64	0.164	NS	NS	NS	NS	NS

Table V. Statistical results of posterior layer thickness in condylar cartilage according to Kruskal-Wallis and Mann Whitney U tests.

Variables	Multiple comparisons between groups										
	Group I Mean ± S.d	Group II Mean ± S.d	Group III Mean ± S.d	Group IV Mean ± S.d	P	I-II	I-III	I-IV	II-III	II-IV	III-IV
Posterior hypertrophic layer (µm)	134.22 ± 90.92	58.89 ± 32.15	59.49 ± 30.25	52.26 ± 34.46	0.015*	0.009**	0.011*	0.007**	NS	NS	NS
Posterior mature layer (µm)	64.80 ± 40.76	31.23 ± 18.04	26.71 ± 19.30	29.27 ± 14.37	0.011*	0.015*	0.007**	0.003**	NS	NS	NS
Posterior proliferative layer (µm)	49.09 ± 32.074	25.34 ± 15.09	22.59 ± 17.21	23.22 ± 12.60	0.021*	0.043*	0.011*	0.003**	NS	NS	NS
Posterior fibrous layer (µm)	66.57 ± 43.54	40.96 ± 19.98	32.86 ± 35.79	49.95 ± 46.10	0.220	NS	NS	NS	NS	NS	NS

Table VI. Statistical results of anterior and posterior total cartilage thickness in condylar cartilage according to Kruskal-Wallis and Mann Whitney U tests.

Variables	Multiple comparisons between groups										
	Group I Mean ± S.d	Group II Mean ± S.d	Group III Mean ± S.d	Group IV Mean ± S.d	P	I-II	I-III	I-IV	II-III	II-IV	III-IV
Anterior total condylar cartilage thickness (µm)	342.198 ± 241.77	212.12 ± 179.79	146.41 ± 75.90	243.38 ± 182.37	0.021*	NS	0.001**	NS	NS	NS	NS
Posterior total condylar cartilage thickness (µm)	314.69 ± 194.15	156.44 ± 70.72	141.66 ± 86.32	154.71 ± 98.83	0.008**	0.009**	0.009**	0.002**	NS	NS	NS

Total condylar cartilage thickness. Histomorphometrical results showed statistically significant difference for anterior total cartilage layer thickness between groups (p<0.05). Group III showed statistically significant low anterior thickness values than group I (control) (p<0.01).

A statistically significant difference was found between the groups for posterior total cartilage layer thickness in condylar cartilage (p<0.01). Experimental groups II (p<0.01), III (p<0.01) and IV (p<0.01) showed statistically significant low thickness values according to group I (control) (Table VI).

DISCUSSION

Our study researched the histomorphometrical effects of different doses LLLT with mandibular advancement appliance on mandibular condyle cartilage of growing rats. The experimental rat model was chosen for this research because the condyle morphology and cartilage layer structures are similar to humans (Oksayan *et al.*, 2015). To investigate experimentally the condylar growth, young and adolescent rats were studied. Similar to our study, Khan *et al.* (2013) studied on 8-week rat condyles for detecting the effects of growth hormone and ultrasound application. Previous experimental rat studies have shown that functional advancement appliances can stimulate and improve mandibular forward position (Rabie *et al.* 2003; Xiong *et al.*; Oksayan *et al.*, 2014). When determining the experimental period we benefited from many other rat mandibular advancement studies, according to these researches the experimental process lasted approximately 4 weeks (Liu *et al.*, 2007; Li *et al.*).

Experimental rat condylar cartilage reaction to biostimulation is now a controversial topic in orthodontic literature. Nowadays, there is scarce research published in orthodontics, regarding the use of LLLT and its impact on biostimulation of condyle cartilage and subchondral bone (Seifi *et al.*; El-Bialy *et al.*, 2015). Laser biostimulation has many effects on cartilage and bone tissues depending on doses.

In this study we revealed that the biostimulatory effects of different LLLT doses (8 J/cm² and 10 J/cm²) on condylar cartilage and growth changes during mandibular advancement. Seifi *et al.* revealed a significant difference between 904 nm LLLT with Diode laser group and control group on rat condyle region, and they concluded that laser irradiation can stimulate condylar growth. Abtahi *et al.* studied the effect of 630 nm LLLT on condylar growth in rabbits and found no increase in cartilage thickness.

The results of our study are in line with Tang & Rabie (2005), they agreed that condylar active growth has a relationship with decreased thickness of cartilage. In addition to this Ghafari & Degroote (1986) found a reduction in total thickness of rat condylar cartilage in mandibular advancement group. Our results referred us that the bite-jumping mandibular advancement appliance 1-2 mm open-bite effect may reduce the TMJ stress and decrease cellular activity in the

proliferative zone and condylar cartilage thickness decreased. Oyonarte *et al.* focused on the ratio between proliferative zone/mature zone and they concluded that reductions could then be shown as an enhancement in proliferative activity of the cartilage.

Histomorphometrical examination showed that subchondral bone area fraction (bony area / total area) was a greater amount in posterior region of condyle in group III but this increase was not statistically significant. Abtahi *et al.* analyzed the thickness of new bone in their LLLT study on rabbits and they found improvement in new bone formation. Oyonarte *et al.* used ultrasound stimulation without mandibular advancement and found increase in subchondral trabecular perimeter by 20 min ultrasound application.

Our results showed that changes are more noticeable in posterior condylar region than anterior part. This situation let us to think on two factors; mesenchymal cell movement and mandibular advancement appliance posterior condylar affect. Rabie *et al.* (2002) support our findings they conclude their results due to an excess of blood vessels, undifferentiated mesenchymal cells reached posterior cartilage region more easily.

According to histomorphometrical findings group III (8 J/cm²) showed more pronounced changes in condylar cartilage layers than group IV (10 J/cm²). Light of Arndt-Schultz law, excess dosage of LLLT can led inhibition on tissue instead of activation (Masuyama *et al.*, 1954).

The power of our study was to have a chance to observe different doses of LLLT and use histomorphometrical analysis for condylar cartilage evaluation. Furthermore, there is scarce information in the literature on mandibular condyle biostimulation with LLLT.

Limitations of this study was the absence of only LLLT groups (ethical committee did not give a permission for naked LLLT groups) without any mandibular advancement appliance, for determining the naked LLLT impact on condylar cartilage tissue.

The clear mechanism through which LLLT can improve condylar growth is not exactly known. These findings may be used as a clinical approach in functional orthopedic therapy, LLLT applications in condylar regions might be useful for shortening functional appliance therapy in skeletal class II patients.

Further studies are needed with more subjects for evaluating the effective LLLT dose on condylar cartilage growth and mandibular advancement

CONCLUSIONS

- Posterior and anterior condylar cartilage layers of rats react differently to varying doses of LLLT and mandibular advancement application. Histomorphometrically posterior condylar cartilage layer changes were found more than anterior region.
- Mandibular advancement application with LLLT produces histomorphometrical changes in condylar cartilage. And maximum changes in condylar cartilage layers were found in 8 J/cm² laser irradiation with mandibular appliance group.
- According to the LLLT impact on condylar cartilage layers, if later studies confirm these results, laser application may be useful in functional orthopedic therapies. And further research is needed to evaluate the different LLLT application parameters and different radiological and cellular analyzing methods.

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OKSAYAN, R.; SÖKÜCÜ, O. & ÜÇÜNCÜ, N. Evaluación histomorfométrica de los efectos del aparato de avance mandibular y la terapia con láser de bajo nivel (TLBN) con diferentes dosis en cartilago condilar y hueso subcondral en ratas. *Int. J. Morphol.*, 38(2):252-258, 2020.

RESUMEN: El objetivo de este estudio fue evaluar los efectos del aparato de avance mandibular y la terapia con láser de bajo nivel (TLBN) con diferentes dosis sobre los cambios hipertróficos celulares, en el cóndilo mandibular de ratas. Cuarenta y ocho ratas albinas macho Wistar de 8 semanas de edad con un peso de 260 y 280 g se dividieron aleatoriamente en cuatro grupos experimentales y control. El grupo I control; grupo II, dispositivos de avance mandibular; grupo III de irradiación con láser de 8 J / cm² (0.25 W, 20 s) con el grupo dispositivos de avance mandibular; y grupo IV con irradiación láser de 10 J / cm² (0,25 W, 25 s) con el grupo de dispositivos de avance mandibular. El cartilago del cóndilo mandibular y los cambios en el hueso subcondral con diferentes dosis de TLBN y dispositivo de avance mandibular, se evaluaron mediante análisis histomorfométrico. Los resultados de la fracción ósea subcondral indicaron que no hubo diferencias significativas entre los grupos (p <0,05). Las diferencias estadísticamente significativas encontradas entre el grupo control y los grupos experimentales, en el grosor del cartilago anterior y posterior (p<0,05) y (p<0,01). Las capas de cartilago condilar posterior y anterior de las ratas reaccionan de ma-

nera diferencial a la aplicación de TLBN y avance mandibular. Se encontraron cambios significativos en las capas de cartilago condilar con irradiación láser de 8 J /cm² con el grupo de dispositivos mandibulares.

PALABRAS CLAVE: Terapia con láser de bajo nivel; Avance mandibular; Rata; Histomorfolométrica.

REFERENCES

- Abtahi, M.; Poosti, M.; Saghravanian, N.; Sadeghi, K. & Shafae, H. The effect of low level laser on condylar growth during mandibular advancement in rabbits. *Head Face Med.*, 8:4, 2012.
- El-Bialy, T.; Alhadlaq, A.; Felemban, N.; Yeung, J.; Ebrahim, A. & Hassan, A. H. The effect of light-emitting diode and laser on mandibular growth in rats. *Angle Orthod.*, 85(2):233-8, 2015.
- El-Bialy, T.; El-Shamy, I. & Graber, T. M. Growth modification of the rabbit mandible using therapeutic ultrasound: is it possible to enhance functional appliance results? *Angle Orthod.*, 73(6):631-9, 2003.
- Ghafari, J. & Degroote, C. Condylar cartilage response to continuous mandibular displacement in the rat. *Angle Orthod.*, 56(1):49-57, 1986.
- Jiao, K.; Dai, J.; Wang, M. Q.; Niu, L. N.; Yu, S. B. & Liu, X. D. Age- and sex-related changes of mandibular condylar cartilage and subchondral bone: a histomorphometric and micro-ct study in rats. *Arch. Oral Biol.*, 55(2):155-63, 2010.
- Khan, I.; El-Kadi, A. O. & El-Bialy, T. Effects of growth hormone and ultrasound on mandibular growth in rats: microct and toxicity analyses. *Arch. Oral Biol.*, 58(9):1217-24, 2013.
- Li, Q.; Zhang, M.; Chen, Y. J.; Zhou, Q.; Wang, Y. J. & Liu, J. Psychological stress alters microstructure of the mandibular condyle in rats. *Physiol. Behav.*, 110-111, 129-39, 2013.
- Liu, C.; Kaneko, S. & Soma, K. Effects of a mandibular lateral shift on the condyle and mandibular bone in growing rats. *Angle Orthod.*, 77(5):787-93, 2007.
- Masuyama, T.; Kobayashi, S. & Otani, A. Influence of ultraviolet irradiation on the growth of bacteria and Arndt-Schultz Law. *Jpn. J. Med. Sci. Biol.*, 7(1):15-23, 1954.
- Oksayan, R.; Sokucu, O. & Ucuncu, N. Effects of bite-jumping appliances on mandibular advancement in growing rats: A radiographic study. *Eur. J. Dent.*, 8(3):291-5, 2014.
- Oksayan, R.; Sokucu, O. & Ucuncu, N. The effects of low-level laser therapy on condylar growth with a mandibular advancement appliance in rats. *Photomed. Laser Surg.*, 33(5):252-7, 2015.
- Owtad, P.; Potres, Z.; Shen, G.; Petocz, P. & Darendeliler, M. A. A histochemical study on condylar cartilage and glenoid fossa during mandibular advancement. *Angle Orthod.*, 81(2):270-6, 2011.
- Oyonarte, R.; Zárate, M. & Rodriguez, F. Low-intensity pulsed ultrasound stimulation of condylar growth in rats. *Angle Orthod.*, 79(5):964-70, 2009.
- Peltomäki, T.; Kylämarkula, S.; Vinkka-Puhakka, H.; Rintala, M.; Kantomaa, T. & Rönning, O. Tissue-separating capacity of growth cartilages. *Eur. J. Orthod.*, 19(5):473-81, 1997.
- Rabie, A. B. M.; She, T. T. & Hägg, U. Functional appliance therapy accelerates and enhances condylar growth. *Am. J. Orthod. Dentofacial Orthop.*, 123(1):40-8, 2003.
- Rabie, A. B. M.; Shum, L. & Chayanupatkul, A. VEGF and bone formation in the glenoid fossa during forward mandibular positioning. *Am. J. Orthod. Dentofacial Orthop.*, 122(2):202-9, 2002.
- Seifi, M.; Maghzi, A.; Gutknecht, N.; Mir, M. & Asna-Ashari, M. The effect of 904 Nm low level laser on condylar growth in rats. *Lasers Med. Sci.*, 25(1):61-5, 2010.

- Tang, G. H. & Rabie, A. B. M. Runx2 regulates endochondral ossification in condyle during mandibular advancement. *J. Dent. Res.*, 84(2):166-71, 2005.
- Tingey, T. F. & Shapiro, P. A. Selective inhibition of condylar growth in the rabbit mandible using intra-articular papain. *Am. J. Orthod.*, 81(6):455-64, 1982.
- Xiong, H.; Hägg, U.; Tang, G. H.; Rabie, A. B. M. & Robinson, W. The effect of continuous bite-jumping in adult rats: a morphological study. *Angle Orthod.*, 74(1):86-92, 2004.

Corresponding author:

Dr. Ridvan Oksayan
Department of Orthodontics
Faculty of Dentistry
Osmangazi University
Eskisehir
TURKEY

Email: ridvan.oksayan@hotmail.com

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