Ultrastructure of the Thyroid Gland in Adult West African Dwarf Goat (Capra hircus)

Ultraestructura de la Glándula Tiroides en la Cabra Enana Africana Occidental (Capra hircus)

Igbokwe Casmir Onwuaso’; Ezeasor Daniel Nwagbo’ & Umar Mohammed Bello”***

SUMMARY: The present study examined ultrastructure of the thyroid gland of adult West African Dwarf (WAD) goat in order to further understand the ultrastructural morphology and some of the changes in the components of the thyroid gland in goat with age. Thyroid glands obtained from fifteen adult WAD goats of different ages and sexes slaughtered at the local abattoirs were used in this study. Electron microscopic techniques were used to study the fixed tissue with emphasis on the follicular and parafollicular cells. The results showed that the ultrastructure is generally similar to that of some domestic animals. Follicular cells were cuboidal in young adult goats thyroids but were flattened in older goat thyroids of 5–7 years. These cells remarkably showed highly dilated cisternae of rough endoplasmic reticulum which decreased in frequency the older goats. Microvilli were short and sparse on the follicular cells and the number decreased in the older goats. Different sizes of apical vesicles of varying electron density were encountered that included colloid droplets, secretory vesicles and lysosome-like bodies and the appearance of these vesicles changed with age. Parafollicular cells were encountered in the basal position between follicular cells in all thyroids examined. Numerous dense cytoplasmic granules were observed and they were not apparently different from that described in several mammals.

KEY WORDS: Ultrastructure; Thyroid gland; Dwarf goat.

INTRODUCTION

The thyroid gland and thyroid hormones are important and central in mammalian development. Some human and animal studies showed that thyroid hormones play a crucial role in nervous, cardiovascular, immune and reproductive system development and function (Janini et al., 1993; Krassas, 2000). In mammalian development, the thyroid hormones have important effects on cell proliferation, differentiation and migration and its effects on growth and metabolism in all stages of mammalian development are well documented. The thyroid generally exhibits similar follicular structure, although there are certain gross, histological and ultrastructural variations amongst the species. Thyroid responses to environmental and nutritional influences do differ amongst domestic animals.

Available information on thyroid structure and development come mainly from invertebrates, murines (mouse, rat), reptiles, zebra fish and humans (Fujita, 1975; 1978; Van Vliet & Polak, 2007; Rupik, 2011). Few published articles on thyroid morphology have considered domestic animals like goat (Roy et al., 1975, 1976; 1978a, 1978b; Baishya et al., 1985, 1986; Osuagwu & Aire, 1992; Adhikary et al., 2003; Bhardwaj et al., 2006). Even so, many of these studies were on gross anatomy, histology and morphometry of thyroid gland of exotic breeds of goats in the temperate and Mediterranean climate. There is no published work on the ultrastructure of the thyroid gland of the West African dwarf (WAD) goat. This breed of goat plays important roles in the diet and socio-economy of the people and traditional livestock farmers in West Africa. The objective of this study is to provide information on the ultrastructural features of the adult thyroid gland of WAD goat with emphasis on the follicular and parafollicular cells.

MATERIAL AND METHOD

Thyroids glands were obtained from fifteen (15) WAD goats of different ages and sexes slaughtered at the local abattoirs in Nsukka and Obollo Afor, Southern Nigeria.

* Department of Veterinary Anatomy, Faculty of Veterinary Medicine, University of Nigeria, Nsukka, Nigeria.
** Department of Anatomy and Physiology, Faculty of Veterinary Science, University of Pretoria, Onderstepoort, South Africa.
*** Department of Veterinary Anatomy, Faculty of Veterinary Medicine, Ahmadu Bello University, Zaria, Nigeria.
The age estimated by dentition (Solaiman, 2010), ranged from 1–7 years.

Immediately after slaughter and excision of the thyroid lobes, small pieces of the organ were diced into 1mm\(^3\) cubes and fixed in 2.5% glutaraldehyde in 0.12 M Millonig’s phosphate buffer at pH 7.4. They were post-fixed in 1% osmium tetroxide after rinsing in phosphate buffer for electron microscopy. The fixed pieces of the thyroid gland were dehydrated in graded ethanol, cleared in propylene oxide and embedded in epoxy resin. Ultra-thin sections (60–80 nm) were collected on copper grids, stained with uranyl acetate, and counterstained with Reynold’s lead citrate and they were examined under Philips CM10 transmission electron microscope accelerating at 80 KV (FEI, Eindhoven, The Netherlands).

RESULTS

The profiles of small and large follicles at low magnification contained colloid in their lumen surrounded by follicular cells that varied in shape and size. Few parafollicular cells were basally located. The follicular cells appeared varied in their lateral and vertical dimensions. Most of these cells were cuboidal in shape. Low columnar cells were rare. Flat follicular cells were present in older thyroids and usually lined the larger follicle, whereas small follicles tend to have cells that were more cuboidal, suggesting therefore that the cell shape may be partly related to the amount of colloid in the lumen of each follicle (Fig. 1). The follicular cells were enclosed by a plasma membrane and the basal part of the membrane was clearly defined without any modifications. The base of the follicular cells and the parafollicular cells rested on a distinct basement membrane. Junctional complexes were quite apparent between the two lateral membranes of follicular cells. It consisted of a tight junction, a gap junction, intermediate junction, desmosomes and septate junction (Fig. 2). The apical regions of the membrane were studded with few stubby microvilli that were apparently more in number in thyroids of 1–2 years than in the older thyroids of 5–7 years. The interfollicular connective tissue spaces contained fine collagen fibrils and fibroblasts with slender processes.
Microvilli were sparse, short and stubby in flat follicular cells of thyroids of 5–7 years. The microvilli on cuboidal cells of adult goats of 1–2 years were more numerous, thinner and distinctly finger-like. The nuclei localized basally, were heterochromatic in most of the cells. It maintained an irregular circular or elliptical shape with some indentations; the nuclei in some younger goats had lobe-like indentations (Fig. 3). It appeared the nuclei shapes were influenced by shape of the cell as well as the various cytoplasmic structures. In the flat cells, therefore the nucleus was flattened, while cuboidal cells contained more rounded nucleus. Nucleoli were prominent and surrounded partly by masses of heterochromatin in the sections examined. The cytoplasm of the follicular cell showed moderate numbers of mitochondria with varied shape that were mostly localized on the apical cytoplasm abutting the colloid and sometimes surrounded by membranes of the rough endoplasmic reticulum (RER) (Fig. 4).

Mitochondria appeared as round, oval, rod-shaped or dumbbell-shaped profiles and varied moderately in shape and size and irregular forms were present. Few free ribosomes were observed and somewhat irregularly distributed in the cytoplasm, some attached but most in small clusters. The cisternae of RER were highly dilated in thyroids of 1–3 years unlike in the older thyroids of 5–7 years. These profiles of RER were more localized in the basal and lateral aspect of the cytoplasm than in the apical cytoplasm. RER were visible as elongated, irregular, elliptical or sometimes circular profiles occupying a considerable portion of the cytoplasm. Cisternae were frequently close to the mitochondria and may occasionally completely surround them. Smooth endoplasmic reticulum was less conspicuous. Golgi complex were well-marked in thyroid sections of all age and consisted of flattened sacs, vacuoles and small vesicles often lied beside or above the nucleus in some sections. The presence of large Golgi complex and colloid droplets commonly found in younger goats (1–2 years) were assumed as evidence of active secretion (Fig. 5). Three types of apically localized granular vesicles were present in the cytoplasm of follicular cells: small round and less dense, large vesicles assumed to becolloid droplets and small round dense bodies that were lysosome-like bodies. The small round and somewhat less dense vesicles were subapically located, while the large colloid droplets were also present with same electron density as the colloid. Small highly electron-dense granules presumed to be primary lysosomes were increased in number in the older thyroids of 5–6 years. In thyroid sections of 1–3 years, large colloid droplets were
frequently observed to have fused with smaller dense granules (probably lysosomes) in sections signifying phagocytosis (pinocytosis) (Fig. 6). However in the older goats of 5–7 years, autophagic vacuoles and heterosomes of various sizes were commonly seen. Some cytoplasmic membrane structures probably multivesicular bodies which were round or irregular in shape were present in the cytoplasm but were more often near the Golgi complex.

Few parafollicular cells (C cells) were generally encountered in sections in the different ages and were positioned basally between two follicular cells, close to the basement membrane and never made contact with the follicular lumen (Fig. 7). Oval to round parafollicular cells were commonly observed, but in some older goats of 5–7 years elongated variety was equally present (Fig. 8). They generally showed oval nuclei, numerous dense secretory granules of varied size and electron density, abundant mitochondrial profile and scant rough endoplasmic reticulum in the cytoplasm. Parafollicular cells appeared to have increased in number of dense secretory granules with age. Lysosome-like bodies, autophagic vacuoles and residual bodies were also observed in these cells with advanced age of 5–7 years (Fig. 9).

DISCUSSION

The general ultrastructural features of the WAD goat thyroid are similar to that of several mammals (Irie, 1960; Fujita, 1975) in terms of the presence of follicular cells, parafollicular cells and their cytoplasmic organelles with minor variations observed in goats. These variations may be due to species and climatic differences. The thyroid of goat possesses features as seen in some domestic animals studied like sheep, camel (Abdel-Magied et al., 2000; Mubarak & Sayed, 2005). These features included the presence of few stunted microvilli, highly dilated cisternae of rough endoplasmic reticulum and abundance of apical vesicles, colloid droplets and lysosomes in the cytoplasm of follicular cells. These organelles varied in number and size with advancing age of the goats as has been observed in some mammals (Fujita, 1975). Few stunted microvilli observed in the younger goat thyroid decreased in the advanced goat of 7 years made of predominantly flattened follicular cells, suggesting decreased endocytic activity. The number and size varies with species and also in experimen-

Fig. 7. EM micrograph of parafollicular cells of thyroid (2 years goat) showing cytoplasm (P) with numerous dense secretory granules (S) and Golgi complex (G). Note the rim of cytoplasm of follicular cell (F) abutting the colloid.

Fig. 8. EM micrograph of thyroid of goat (6 years) showing elongated parafollicular cells with numerous polarized dense granules amongst other organelles. Note thin rim of cytoplasm of follicular cells (F) close to follicular colloid (C).

Fig. 9. EM micrograph of parafollicular cell showing numerous organelles in the cytoplasm that included mitochondria (M), dense secretory granules (S), lysosomes (L). Note the thin rim of follicular cell (F) cytoplasm abutting the colloid lumen (C).

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PALABRAS CLAVE: Ultraestructura; Glandula tiroides; Cabra enana.
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Correspondence to: Igbokwe Casimir Onwuaso
Senior Lecturer
Department of Veterinary Anatomy
Faculty of Veterinary Medicine University of Nigeria
Nsukka
NIGERIA
Email: casmir.igbokwe@unn.edu.ng

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